

## Helicobacter Pylori Stain Kit- Technical Memo

### KIT INCLUDES:

Solution A: Periodic Acid 1%, Aqueous	250 ml	<b>Part 9130A</b>
Solution B: Sodium Metabisulfite 5%, Acidified	250 ml	
Solution C: Alcian Yellow Stain 1%, Alcoholic	250 ml	
Solution D: Toluidine Blue Stock Stain 1%, Aqueous	10 ml	
Solution E: Sodium Hydroxide 3%, Aqueous	10 ml	

**COMPLIMENTARY POSITIVE CONTROL SLIDES:** Enclosed are two complimentary unstained positive control slides for the initial verification of staining techniques and reagents. Verification must be documented by running one Newcomer Supply complimentary positive control slide along with your current positive control slide for the first run. Retain the second complimentary control slide for further troubleshooting, if needed.

*Individual stain solutions and control slides may be available for purchase under separate part numbers at [www.newcomersupply.com](http://www.newcomersupply.com).*

### Additionally Needed:

Xylene, ACS	Part 1445
Alcohol, Ethyl Denatured, 100%	Part 10841
Alcohol, Ethyl Denatured, 95%	Part 10842

**For storage requirements and expiration date refer to individual bottle labels.**

### APPLICATION:

Newcomer Supply Helicobacter Pylori Stain Kit procedure is used for the detection of *Helicobacter pylori* (*H. pylori*) in gastrointestinal specimens.

### METHOD:

**Fixation:** Formalin 10%, Phosphate Buffered (Part 1090)

**Technique:** Paraffin sections cut at 4 microns

**Solutions:** All solutions are manufactured by Newcomer Supply, Inc.

All Newcomer Supply Stain Kits are designed to be used with Coplin jars filled to 40 ml following the provided staining procedure. Some solutions in the kit may contain extra volumes.

### STAINING PROCEDURE:

- If necessary, heat dry tissue sections/slides in oven.
- Deparaffinize sections thoroughly in three changes of xylene, 3 minutes each. Hydrate through two changes each of 100% and 95% ethyl alcohols, 10 dips each. Wash well with distilled water.
  - See Procedure Note #1.
- Oxidize in Solution A: Periodic Acid 1%, Aqueous; 10 minutes.
- Wash slides well with tap water.
- Place in Solution B: Sodium Metabisulfite 5%, Acidified; 5 minutes.
- Wash in tap water for 5 minutes.
- Stain in Solution C: Alcian Yellow Stain 1%, Alcoholic; 5 minutes.
- Wash well in tap water.
- Prepare fresh Toluidine Blue Working Solution; combine, mix well.
  - Distilled Water 50 ml
  - Solution D: Toluidine Blue Stock Stain 1%, Aqueous 0.5 ml
  - Solution E: Sodium Hydroxide 3%, Aqueous 2 drops
- Stain in fresh Toluidine Blue Working Solution; 3 minutes.
- Wash well in tap water.
- Allow slides to air-dry.
- Clear dried slides in three changes of xylene, 10 dips each; coverslip with compatible mounting medium.
  - Eliminate alcohol dehydration to retain organism staining.

### RESULTS:

<i>Helicobacter pylori</i>	Blue
Mucin	Yellow
Background	Pale blue

### PROCEDURE NOTES:

- Drain slides after each step to prevent solution carry over.
- If using a xylene substitute, follow manufacturer recommendations for deparaffinization and clearing steps.

### REFERENCES:

- Carson, Freida L., and Christa Hladik Cappellano. *Histotechnology: A Self-instructional Text*. 4th ed. Chicago: ASCP Press, 2015. 225-226.
- Leung, Jennifer, Kevin Gibbon and Robert Vartaniam. "Rapid Staining Method for *Helicobacter pylori* in Gastric Biopsies." *The Journal of Histotechnology* 19.12 (1996): 131-132.
- Modifications developed by Newcomer Supply Laboratory.