

Fungus, Grocott Methenamine Silver (GMS) Stain Kit - Technical Memo

KIT INCLUDES:

	Part 9121A	Part 9121B
Solution A: Chromic Acid 5%, Aqueous	250 ml	500 ml
Solution B: Sodium Bisulfite 1%, Aqueous	250 ml	500 ml
Solution C: Silver Nitrate	125 ml	250 ml
Solution D: Methenamine Borate	125 ml	250 ml
Solution E: Gold Chloride 0.1%, Aqueous	250 ml	500 ml
Solution F: Sodium Thiosulfate 2%, Aqueous	250 ml	500 ml
Solution G: Light Green SF Yellowish Stain 0.2%, Aqueous	250 ml	500 ml

COMPLIMENTARY POSITIVE CONTROL SLIDES: Enclosed are two complimentary unstained positive control slides for the initial verification of staining techniques and reagents. Verification must be documented by running one Newcomer Supply complimentary positive control slide along with your current positive control slide for the first run. Retain the second complimentary control slide for further troubleshooting, if needed.

Individual stain solutions and additional control slides may be available for purchase under separate part numbers at www.newcomersupply.com.

Additionally Needed:

Hydrochloric Acid 5%, Aqueous	Part 12086 (for acid cleaning glassware)
Xylene, ACS	Part 1445
Alcohol, Ethyl Denatured, 100%	Part 10841
Alcohol, Ethyl Denatured, 95%	Part 10842
Coplin Jar, Plastic	Part 5184 (for microwave modification)

For storage requirements and expiration date refer to individual bottle labels.

APPLICATION:

Newcomer Supply Fungus, Grocott Methenamine Silver (GMS) Stain Kit procedure, with included microwave modification, is one of the best staining methods to demonstrate a variety of fungal organisms including: *Pneumocystis*, *Aspergillus*, *Blastomyces*, *Candida* and *Histoplasma*.

METHOD:

Fixation: Formalin 10%, Phosphate Buffered (Part 1090)

Technique: Paraffin sections cut at 4 microns.

Solutions: All solutions are manufactured by Newcomer Supply, Inc.

All Newcomer Supply Stain Kits are designed to be used with Coplin jars filled to 40 ml following the staining procedure provided below. Some solutions in the kit may contain extra volumes.

PRESTAINING PREPARATION:

- If necessary, heat dry tissue sections/slides in oven.
- All glassware/plasticware must be acid cleaned prior to use.
 - See Procedure Notes #1 and #2 (page 2).
- Prepare Silver-Methenamine Working Solution and mix well.
 - Solution C: Silver Nitrate 20 ml
 - Solution D: Methenamine Borate 20 ml
- Preheat Silver-Methenamine Working Solution to 45°C-60°C in a water bath approximately 20 to 30 minutes before use.
 - See Procedure Note #3 (page 2).
 - Do not preheat if using Microwave Modification; Step 11.**

STAINING PROCEDURE:

- Deparaffinize sections thoroughly in three changes of xylene, 3 minutes each. Hydrate through two changes each of 100% and 95% ethyl alcohols, 10 dips each. Wash well with distilled water.
 - See Procedure Notes #4 and #5 (page 2).
- Oxidize in Solution A: Chromic Acid 5%, Aqueous for 1 hour.

Microwave Modification: See Procedure Note #6 (page 2)

 - Oxidize slides in a plastic Coplin jar containing Solution A: Chromic Acid 5%, Aqueous and microwave for 1 minute and 20 seconds at 60°C.
- Wash well in running tap water; rinse in distilled water.

- Place in Solution B: Sodium Bisulfite 1%, Aqueous for 1 minute.
- Wash for 5 minutes in running tap water; rinse well in distilled water.
- Incubate slides in preheated Silver-Methenamine Working Solution (Step #4) at 45°C-60°C or at room temperature, for 12-18 minutes until sections appear paper-bag brown.
 - Periodically remove control, rinse in warm distilled water, check microscopically for adequate silver impregnation. Fungi should be dark brown.
 - If organisms are not sufficiently dark, return slides to warm silver solution. Recheck at 2-3 minute intervals until desired intensity is achieved.
 - Pneumocystis* may take longer to stain than other fungus.
 - Staining at room temperature will require longer incubation.
- Microwave Modification:**
 - Incubate slides in a plastic Coplin jar containing Silver-Methenamine Working Solution and microwave for 1 minute at 70°C.
 - Check microscopically for adequate development.
 - If additional incubation is required, return slides to warm silver solution. Recheck at 2-3 minute intervals.
- Rinse in three to four changes of distilled water.
 - Never use tap water at this step.
- Tone in Solution E: Gold Chloride 0.1%, Aqueous until sections turn gray; 20 seconds to 1 minute.
- Rinse well in distilled water.
- Remove unreduced silver in Solution F: Sodium Thiosulfate 2%, Aqueous for 2 minutes.
- Wash in running tap water for 5 minutes; rinse in distilled water.
- Counterstain in Solution G: Light Green SF Yellowish Stain 0.2%, Aqueous for 2 minutes.
- Dehydrate quickly in two changes each of 95% and 100% ethyl alcohol. Clear in three changes of xylene, 10 dips each; coverslip with compatible mounting medium.

RESULTS:

Fungi	Crisp black cell walls with visible internal structures
Background	Green
Mucin	Taupe to dark gray

PROCEDURE NOTES:

1. Acid clean all glassware/plasticware (12086) and rinse thoroughly in several changes of distilled water.
2. Plastic (5500), plastic-tipped or paraffin coated metal forceps must be used with any silver solution to prevent precipitation of silver salts. No metals of any kind should be in contact with any silver solution. Only glass thermometers should be used.
3. Staining at higher temperatures means faster development but may cause precipitate to form in the working silver solution and deposit on slides. Maintaining silver solution between 45°C-60°C will minimize precipitate.
4. Drain slides after each step to prevent solution carry over.
5. Do not allow sections to dry out at any point during procedure.
6. The suggested microwave procedure has been tested at Newcomer Supply. This procedure is a guideline and techniques should be developed for use in your laboratory.
7. If using a xylene substitute, closely follow the manufacturer's recommendations for deparaffinization and clearing steps.

REFERENCES:

1. Carson, Freida L., and Christa Hladik. *Histotechnology: A Self-Instructional Text*. 3rd ed. Chicago, Ill.: American Society of Clinical Pathologists, 2009. 239-243.
2. Grocott, R G, "A Stain for Fungi in Tissue Sections and Smears using Gomori Methenamine Silver Nitrate Technic". *American Journal of Clinical Pathology* 25 (1955): 975-979.
3. Koski, John. "Silver Methenamine Borate (SMB): Cost Reduction with Technical Improvement in Silver Nitrate-Gold Chloride Impregnations." *The Journal of Histotechnology* 4.3 (1981): 115-119.
4. Sheehan, Dezna C., and Barbara B. Hrapchak. *Theory and Practice of Histotechnology*. 2nd ed. St. Louis: Mosby, 1980. 245-246.
5. Modifications developed by Newcomer Supply Laboratory.