

Wright Stain, Modified for Smears - Technical Memo

SOLUTION:	500 ml	1 Liter	1 Gallon
Wright Stain, Modified	Part 1421A	Part 1421B	Part 1421C

<u>Additionally Needed:</u>	
Alcohol, Methanol Anhydrous, ACS	Part 12236
Wright Stain Buffer, pH 6.8	Part 1430

For storage requirements and expiration date refer to individual bottle labels.

APPLICATION:

Newcomer Supply Wright Stain, Modified for Smears, provides a concentrated Wright's formula for differential staining of cell types in peripheral blood smears and bone marrow smears/films. This procedure is applicable for either hand or automated staining processes.

METHOD:

Technique: Flat staining rack method

Solutions: All solutions are manufactured by Newcomer Supply, Inc.

PRESTAINING PREPARATION:

1. Prepare within an accepted time frame, a well-made blood smear or bone marrow smear/film per your laboratories protocol, with a focus on uniform cell distribution.
2. Allow slides to thoroughly air-dry prior to staining.
3. Filter Wright Stain, Modified prior to use with quality filter paper.
 - a. *Filter sufficient stain to allow 1 ml of stain per slide.*

STAINING PROCEDURE:

4. Place slides on flat staining rack suspended over sink.
5. Fix by flooding slides with Methanol (12236); 10-30 seconds.
6. Drain off Methanol.
7. Flood each slide with 1 ml of filtered Wright Stain, Modified for 3-5 minutes.
 - a. *See Procedure Notes #1 and #2.*
8. Retain Wright Stain, Modified on slides.
9. Directly add 1 ml of Wright Stain Buffer, pH 6.8 (1430) to each slide; agitate gently to mix with retained Wright Stain.
10. Stain for an additional 6-10 minutes.
11. Wash well in distilled water; rinse thoroughly in running tap water.
12. Air-dry slides in a vertical position; examine microscopically.
13. If coverslip is preferred, allow slides to air-dry and coverslip with compatible mounting medium.

RESULTS:

Erythrocytes	Pink
Neutrophils	Granules - Purple
Eosinophils	Granules - Pink
White blood cells	Chromatin - Purple
Lymphocytes	Cytoplasm - Blue
Monocytes	Cytoplasm- Blue
Bacteria	Deep Blue

PROCEDURE NOTES:

1. Timings provided are suggested ranges. Optimal times will depend upon staining intensity preference.
2. Smears containing primarily normal cell populations require minimum staining time; immature cells and bone marrow smears/films may require longer staining times.
3. The color range of stained cells may vary depending on buffer pH and pH of rinse water.
 - a. *Alkalinity is indicated by red blood cells being blue-grey and white blood cells only blue.*
 - b. *Acidity is indicated by red blood cells being bright red or pink and lack of proper staining in white blood cells.*
 - c. *If necessary, adjust buffer pH accordingly to 6.8 +/- 0.2.*

REFERENCES:

1. Lillie, R. D., and Harold Fullmer. *Histopathologic Technic and Practical Histochemistry*. 4th ed. New York: McGraw-Hill, 1976. 747-748.
2. McPherson, Richard and Matthew Pincus. *Henry's Clinical Diagnosis and Management by Laboratory Methods*. 22nd ed. Philadelphia: Elsevier Saunders, 2011. 522-532.
3. Sheehan, Dezna C., and Barbara B. Hrapchak. *Theory and Practice of Histotechnology*. 2nd ed. St. Louis: Mosby, 1980. 154-155.
4. Modifications developed by Newcomer Supply Laboratory.

Wright-Giemsa Stain, Modified for Tissue Sections - Technical Memo

SOLUTIONS:	500 ml	1 Liter	1 Gallon
Wright Stain, Modified	Part 1421A	Part 1421B	Part 1421C
Giemsa Stock Stain	Part 1120A	Part 1120B	

Additionally Needed:

Giemsa Control Slides	Part 4240
Xylene, ACS	Part 1445
Alcohol, Ethyl Denatured, 100%	Part 10841
Alcohol, Ethyl Denatured, 95%	Part 10842
Alcohol, Methanol Anhydrous, ACS	Part 12236

For storage requirements and expiration date refer to individual product labels.

APPLICATION:

Newcomer Supply Wright-Giemsa Stain, Modified for Tissue Sections combines a modified Wright's formula with a Giemsa Stain Solution for differential staining of hematopoietic tissue and demonstration of bacteria that may be present in the sections. This procedure is applicable for either hand or automated staining processes.

METHOD:

- Fixation:** Recommended for hematopoietic tissue:
- Zenker Fixative, Modified, Zinc Chloride (Part 1461)*
 - B-5 Fixative Modified, Zinc Chloride (Part 1015)*
 - Formalin 10%, Phosphate Buffered (Part 1090)*

Technique: Paraffin sections cut at 4 microns

Solutions: All solutions are manufactured by Newcomer Supply, Inc.

All Newcomer Supply stain procedures are designed to be used with Coplin jars filled to 40 ml following the provided staining procedure.

PRESTAINING PREPARATION:

- If necessary, heat dry tissue sections/slides in oven.
- Prepare **fresh** Working Giemsa Stain:

a. Distilled Water	40 ml
b. Giemsa Stock Stain (1120)	5 ml

STAINING PROCEDURE:

- Deparaffinize sections thoroughly in three changes of xylene, 3 minutes each. Hydrate through two changes each of 100% and 95% ethyl alcohols, 10 dips each. **Stop at 95% ethyl alcohol.**
 - See Procedure Notes #1 and #2.
- Treat slides in two changes of Methanol (12236); 3 minutes each.
- Stain in Wright Stain, Modified for 6 minutes.
- Stain in **fresh** Working Giemsa Stain (Step #2); 60°C oven for 60 minutes.
- Dehydrate in two changes each of 95% and 100% ethyl alcohol. Clear in three changes of xylene, 10 dips each; coverslip with compatible mounting medium.

RESULTS:

Nuclei	Blue
Cytoplasm	Pink to red
Bacteria	Blue

PROCEDURE NOTES:

- Drain slides after each step to prevent solution carry over.
- Do not allow sections to dry out at any point during procedure.
- The color range of stained cells may vary depending upon fixation.
- If using a xylene substitute, closely follow the manufacturer's recommendations for deparaffinization and clearing steps.

REFERENCES:

- Shapiro, Stanley H., and Hilda Laufer. "Observations on Fixation and Staining of Bone Marrow Biopsies." *The Journal of Histotechnology* 11.3 (1988): 145-47.
- Sheehan, Dezna C., and Barbara B. Hrapchak. *Theory and Practice of Histotechnology*. 2nd ed. St. Louis: Mosby, 1980. 155-156.
- Modifications developed by Newcomer Supply Laboratory.