

Newly Blue Stain Set - Technical Memo

SET INCLUDES: **Part 1258A**

Solution A: Celestine Blue Stain 1%, Aqueous 250ml
Solution B: Ferric Ammonium Sulfate 4%, Aqueous 250 ml

Additionally Needed For H&E Staining:

Hematoxylin and Eosin (H&E) Control Slides	Part 4278		
Xylene, ACS	Part 1445		
Alcohol, Ethyl Denatured, 100%	Part 10841		
Alcohol, Ethyl Denatured, 95%	Part 10842		
Acid Alcohol 1%	Part 10011		
Lithium Carbonate, Saturated Aqueous	Part 12215	or	Scott Tap Water Substitute Part 1380
Alcohol, Ethyl Denatured, 70%	Part 10844		
Eosin Y Working Solution	Part 1072		

For storage requirements and expiration date refer to individual product labels.

APPLICATION:

Newcomer Supply Newly Blue Stain Set provides a synthetic nuclear stain (hematoxylin substitute), that is indistinguishable from standard hematoxylin staining results. Newly Blue nuclear staining is crisp with well delineated purple to blue nuclei and displays a clear contrast to cytoplasmic stains for precise cellular interpretation.

METHOD:

Fixation: Formalin 10%, Phosphate Buffered (Part 1090)
Technique: Paraffin sections cut at 4 microns
Solutions: All solutions are manufactured by Newcomer Supply, Inc.

All Newcomer Supply Staining Sets are designed to be used with Coplin jar filled to 40 ml following the provided staining procedure.

PRESTAINING PREPARATION:

1. If necessary, heat dry tissue sections/slides in oven.
2. Prepare Newly Blue Working Solution and mix well:
 - a. *Solution A: Celestine Blue Stain 1%, Aqueous* 20 ml
 - b. *Solution B: Ferric Ammonium Sulfate 4%, Aqueous* 20 ml
 - c. *Filter before use.*
 - d. *See Procedure Notes #1 & #2.*

STAINING PROCEDURE:

3. Deparaffinize sections thoroughly in three changes of xylene, 3 minutes each. Hydrate through two changes each of 100% and 95% ethyl alcohols, 10 dips each. Wash well with distilled water.
 - a. *See Procedure Notes #3 and #4.*
4. Stain with Newly Blue Working Solution for 4 minutes.
5. Wash well in three changes of tap water.
6. Differentiate quickly in Acid Alcohol 1% (10011).
 - a. *Nuclei should be distinct; background light to colorless.*
7. Rinse well in three changes of tap water.
8. Blue in Lithium Carbonate, Saturated Aqueous (12215) or Scott Tap Water Substitute (1380) for 10 dips.
9. Wash in three changes of tap water; rinse in distilled water.
10. Drain excess water; proceed to 70% alcohol for 10 dips.
11. Counterstain in Eosin Y Working Solution (1072) or prepared Eosin-Phloxine Working Solution (1082) for 30 seconds to 3 minutes, depending on preference of intensity.
12. Dehydrate in two changes of 95% ethyl alcohol for 1 minute each and two changes of 100% ethyl alcohol, 10 dips each. Clear in three changes of xylene, 10 dips each; coverslip with compatible mounting medium.

RESULTS:

Nuclei Blue
Cytoplasm and other tissue elements Various shades of pink

PROCEDURE NOTES:

1. Newly Blue Working Solution is stable for up to 5 days.
2. Blot off any surface sheen that may develop prior to use.
3. Drain slides after each step to prevent solution carry over.
4. Do not allow sections to dry out at any point during procedure.
5. If using a xylene substitute, closely follow the manufacturer's recommendations for deparaffinization and clearing steps.

REFERENCES:

1. Carson, Freida L., and Christa Hladik Cappellano. *Histotechnology: A Self-instructional Text*. 4th ed. Chicago: ASCP Press, 2015. 115-116.
2. Sheehan, Dezna C., and Barbara B. Hrapchak. *Theory and Practice of Histotechnology*. 2nd ed. St. Louis: Mosby, 1980. 148-150.
3. Modifications developed by Newcomer Supply Laboratory.