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Part 1905 May 2020

# Alpha Amylase 1%, Aqueous for Glycogen Digestion - Technical Memo

SOLUTION: 500 ml
Alpha Amylase 1%, Aqueous Part 1905B

**Additionally Needed:** 

Periodic Acid Schiff (PAS) Glycogen Control Slides Part 4540 Periodic Acid 0.5%, Aqueous Part 13308 Part 1371 Schiff Reagent, McManus Hematoxylin Stain, Harris Part 12013 Acid Alcohol 1% Part 10011 Lithium Carbonate, Saturated Aqueous Part 12215 Xylene, ACS Part 1445 Alcohol, Ethyl Denatured, 100% Part 10841 Alcohol, Ethyl Denatured, 95% Part 10842

Coplin Jar, Plastic Part 5184 (for glycogen digestion microwave modification)

For storage requirements and expiration date refer to individual product labels.

#### **APPLICATION:**

Newcomer Supply Alpha Amylase 1%, Aqueous is a convenient readyto-use glycogen digestion solution for aiding in further identification of mucosubstances used in conjunction with the Periodic Acid Schiff (PAS) Stain procedure.

### **METHOD:**

Fixation: Formalin 10%, Phosphate Buffered (Part 1090)

Technique: Paraffin sections cut at 4 microns

## **STAINING PROCEDURE:**

- 1. If necessary, heat dry tissue sections/slides in oven.
- Deparaffinize sections thoroughly in three changes of xylene, 3
  minutes each. Hydrate through two changes each of 100% and
  95% ethyl alcohols, 10 dips each. Wash well with distilled water.
  - a. See Procedure Notes #1 and #2.
- Digestion Step: See Procedure Note #3.
  - a. Two control slides and two patient slides are needed.
  - b. Label one control slide and one patient slide "with".
  - c. Label the other control slide and patient slide "without".
  - d. Place slides labeled "without" in separate Coplin jar of distilled water; hold for Step #5.
  - e. Apply Alpha Amylase 1%, Aqueous to slides labeled "with" for 30 minutes at room temperature.
  - f. Proceed to Step #5.
- 4. Digestion Microwave Modification: See Procedure Note #4.
  - a. Follow Steps #3a through #3d.
  - b. Place slides labeled "with" in a <u>plastic</u> Coplin jar containing Alpha Amylase 1%, Aqueous and microwave for 1 minute at 37°C. Let sit in warm solution for an additional minute.
- Combine all slides for remaining steps; wash in running tap water for 1 minute, rinse in distilled water.
- 6. Place in Periodic Acid 0.5%, Aqueous (13308) for 10 minutes.
- 7. Wash in three changes of tap water; rinse in distilled water.
- 8. Place in Schiff Reagent, McManus (1371) for 20 minutes.
- 9. Wash in lukewarm tap water for 5 minutes.
- Stain with Hematoxylin Stain, Harris (12013), 1-5 minutes, depending on preference of nuclear stain intensity.
- 11. Wash in tap water for 2-3 minutes.
- 12. Differentiate in Acid Alcohol 1% (10011); 1-2 quick dips.
- Wash in tap water for 1 minute.

- 14. Blue in Lithium Carbonate, Saturated Aqueous (12215); 3-4 dips.
- 15. Wash in several changes of tap water; rinse in distilled water.
- Dehydrate in two changes each of 95% and 100% ethyl alcohol.
   Clear in three changes of xylene, 10 dips each; coverslip with compatible mounting medium.

#### RESULTS:

Glycogen Magenta

Glycogen digestion Absence of magenta

Acid & neutral epithelial mucin Magenta Nuclei Blue

## **PROCEDURE NOTES:**

- 1. Drain slides after each step to prevent solution carry over.
- 2. Do not allow sections to dry out at any point during procedure.
- 3. Slides labeled "with" will be treated with amylase digestion, slides labeled "without" will not be treated for digestion.
- The suggested microwave procedure has been tested at Newcomer Supply. This procedure is a guideline and techniques should be developed for use in your laboratory.
- If using a xylene substitute, closely follow the manufacturer's recommendations for deparaffinization and clearing steps.

### **REFERENCES:**

- Bancroft, John D., and Marilyn Gamble. Theory and Practice of Histological Techniques. 6th ed. Oxford: Churchill Livingstone Elsevier, 2008. 168-171, 180.
- Carson, Freida L., and Christa Hladik. Histotechnology: A Self-Instructional Text. 3rd ed. Chicago, Ill.: American Society of Clinical Pathologists, 2009.137-141.
- Sheehan, Dezna C., and Barbara B.Hrapchak. Theory and Practice of Histotechnology. 2nd ed. St. Louis: Mosby, 1980. 164-168.
- 4. Modifications developed by Newcomer Supply Laboratory.

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