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Part 3188 Revised January 2020

# **CD138 Control Slides – Technical Memo**

#### CONTROL SLIDES:

Part 3188A 10 Slide/Set Part 3188B 98 Slide/Set

## PRODUCT SPECIFICATIONS:

**Tissue:** Positive staining tonsil and negative staining adipose.

Fixation: Formalin 10%, Phosphate Buffered (Part 1090).

Section/Glass: Paraffin sections cut at 4 microns on Superfrost<sup>™</sup> Plus slides.

Quality Control Stain: CD138 quality control stained slide(s) included.

**Reactivity:** Guaranteed product specific reactivity for one year from date of receipt. Revalidate after one year to verify continued reactivity. **Storage:** 15-30°C in a light deprived and humidity controlled environment.

Intended Use: To verify histological techniques and reagent reactivity.

Before using unstained control slides, review the enclosed stained slide(s) to ensure that this tissue source is acceptable for testing needs.

#### **APPLICATION:**

Newcomer Supply CD138 Control Slides are for the positive immunohistochemical staining of CD138, a specific marker for identification of plasma cells and plasma cell differentiation within hematolymphoid tissues in benign and neoplastic conditions.

#### NEWCOMER SUPPLY VALIDATION PROCEDURE:

- 1. Heat dry sections in oven according to your laboratory protocol.
- Deparaffinize sections thoroughly in three changes of xylene, 3 minutes each. Hydrate through two changes each of 100% and 95% ethyl alcohols, 10 dips each. Wash well with distilled water.
  a. See Procedure Note #1.
- 3. Proceed, if necessary, with an epitope/antigen retrieval technique approved for use in your laboratory.
- 4. Rinse in distilled water; tap off excess water.
- 5. Circle sections with Pap Pen Liquid Blocker (Part 6505, 6506 or 6507) to reduce reagent usage and ensure tissue coverage.
- Block endogenous peroxidase with freshly made 3% Hydrogen Peroxide. Incubate for 5 minutes.
  - a. See Procedure Note #2.
- 7. Wash slides gently in distilled water. Rinse in two changes of Tris Buffered Saline.
  - a. See Procedure Note #3.
- 8. Tap off excess buffer; apply CD138 primary antibody. Incubate at room temperature for 30 minutes.
- 9. Rinse slides in two changes of buffer.
- 10. Tap off excess buffer; apply HRP-Polymer solution. Incubate for 20 minutes.
- 11. Rinse slides in two changes of buffer.
- 12. Prepare required quantity of DAB substrate/chromogen.
- 13. Tap off excess buffer; apply DAB. Incubate for 5 minutes.
- 14. Rinse slides in four changes of distilled water.
- 15. Counterstain lightly with Hematoxylin Stain, Gill I (Part 1180) for 5 minutes.
- 16. Rinse slides in warm tap water to blue sections.
- 17. Dehydrate in two changes each of 95% and 100% ethyl alcohol. Clear in three changes of xylene, 10 dips each; coverslip with compatible mounting medium.

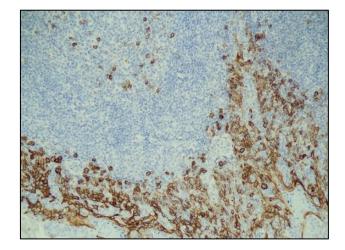
#### **RESULTS:**

## PROCEDURE NOTES:

- 1. Do not allow sections to dry out at any point during procedure.
- 2. Dilute sufficient Hydrogen Peroxide 30%, Aqueous (Part 1206) with distilled water to a 3% (1/10) solution prior to use.
- Dilute sufficient Tris Buffered Saline 0.05M, pH 7.6, 10X (Part 140304) with distilled water to a 1/10 solution prior to use for all buffer rinses in this procedure.
- Biocare CD138 (B-A38) is the concentrated primary antibody used. Dilute primary antibody to 1/100 working dilution with Biocare Van Gogh Yellow Diluent (PD902).
- 5. Biocare Mach 2<sup>™</sup> Universal HRP-Polymer Detection (M2U522) is the HRP-Polymer solution used.
- 6. Cell Marque DAB Substrate Kit (957D) is the chromogen used.
- 7. If using a xylene substitute, closely follow the manufacturer's recommendations for deparaffinization and clearing steps.

#### **REFERENCES:**

- 1. Biocare CD138 Antibody datasheet.
- 2. Biocare Van Gogh Yellow Diluent datasheet.
- 3. Biocare Mach 2<sup>™</sup> Universal HRP-Polymer Detection datasheet.
- 4. Cell Marque DAB Substrate Kit datasheet.
- 5. Modifications developed by Newcomer Supply Laboratory.



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