

Pavi

Part 4695 Revised January 2020

Trichrome, Uterus Control Slides - Technical Memo

CONTROL SLIDES: Part 4695A Part 4695B 10 Slide/Set 98 Slide/Set

PRODUCT SPECIFICATIONS:

Tissue: Positive staining uterus.

Fixation: Formalin 10%. Phosphate Buffered (Part 1090).

Section/Glass: Paraffin sections cut at 4 microns on Superfrost™ Plus slides.

Quality Control Stain: Gomori One-Step Aniline Blue quality control stained slide(s) included.

Reactivity: Guaranteed product specific reactivity for one year from date of receipt. Revalidate after one year to verify continued reactivity.

Storage: 15-30°C in a light deprived and humidity controlled environment. **Intended Use:** To verify histological techniques and reagent reactivity.

Before using unstained control slides, review the enclosed stained slide(s) to ensure that this tissue source is acceptable for testing needs.

CONTROL SLIDE VALIDATION:

With Trichrome, Gomori One-Step, Aniline Blue Stain Kit:	Part 9176B/A	Individual Stain Solution
Solution A: Bouin Fluid	250/500 ml	Part 1020
Solution B: Ferric Chloride, Acidified	125/250 ml	Part 1409
Solution C: Hematoxylin 1%, Alcoholic	125/250 ml	Part 1409
Solution D: Trichrome Stain, Gomori One-Step, Aniline Blue	250/500 ml	Part 1403
Solution E: Acetic Acid 0.5%, Aqueous	250/500 ml	Part 100121

APPLICATION:

Newcomer Supply Trichrome, Uterus Control Slides are for the positive histochemical staining of connective tissue and to differentially demonstrate collagen and muscle fibers.

PRESTAINING PREPARATION:

- 1. Heat dry sections in oven according to your laboratory protocol.
- 2. Preheat Solution A: Bouin Fluid to 56-60°C in oven or water bath. (Skip if using overnight method or microwave procedure.)

NEWCOMER SUPPLY VALIDATION PROCEDURE:

- Deparaffinize sections thoroughly in three changes of xylene, 3 minutes each. Hydrate through two changes each of 100% and 95% ethyl alcohols, 10 dips each. Wash well with distilled water.
 - a. See Procedure Notes #1 and #2.
- Mordant in preheated Solution A: Bouin Fluid (Step #2) for one hour at 56-60°C or overnight at room temperature. Cool at room temperature for 5-10 minutes.
 - a. Skip Step #4 if tissue was originally Bouin fixed.

Microwave Modification: See Procedure Note #3.

- b. Place slides in a <u>plastic</u> Coplin jar containing Solution A: Bouin Fluid and microwave for 5 minutes at 60°C.
- 5. Wash well in running tap water; rinse in distilled water.
- 6. Prepare fresh Weigert Iron Hematoxylin; combine and mix well.
 - a. Solution B: Ferric Chloride, Acidified 20 ml
 - b. Solution C: Hematoxylin 1%, Alcoholic 20 ml
- 7. Stain in fresh Weigert Iron Hematoxylin for 10 minutes.
- Wash in running tap water for 10 minutes; rinse in distilled water.
 a. See Procedure Note #4..
- Stain with Solution D: Trichrome Stain, Gomori One-Step, Aniline Blue for 20 minutes.
- 10. Differentiate in Solution E: Acetic Acid 0.5%, Aqueous; 2 minutes.
- 11. Rinse quickly in distilled water.
- Dehydrate in two changes each of 95% and 100% ethyl alcohol. Clear in three changes of xylene, 10 dips each; coverslip with compatible mounting medium.

RESULTS:

Collagen and mucin
Muscle fibers, cytoplasm and keratin
Nuclei

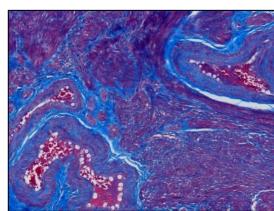
Blue
Red
Blue/black

PROCEDURE NOTES:

- 1. Drain slides after each step to prevent solution carry over.
- 2. Do not allow sections to dry out at any point during procedure.
- The suggested microwave procedure has been tested at Newcomer Supply. This procedure is a guideline and techniques should be developed for use in your laboratory.
- If Weigert Iron Hematoxylin is not completely washed from tissue sections, nuclear and cytoplasmic staining may be compromised.
- Trichrome, Uterus Control Slides are validated with Trichrome Stain Kit, Gomori One-Step, Aniline Blue but can be used as positive controls with any preferred Trichrome procedure.
- 6. If using a xylene substitute, closely follow the manufacturer's recommendations for deparaffinization and clearing steps.

REFERENCES:

- Brown, Richard. Histologic Preparations: Common Problems and Their Solutions. Northfield, Ill.: College of American Pathologists, 2009. 95-101.
- Carson, Freida L., and Christa Hladik. Histotechnology: A Self-Instructional Text. 3rd ed. Chicago, Ill.: American Society of Clinical Pathologists, 2009. 165-166.
- Sheehan, Dezna C., and Barbara B. Hrapchak. Theory and Practice of Histotechnology. 2nd ed. St. Louis: Mosby, 1980. 191-192
- Vacca, Linda L. Laboratory Manual of Histochemistry. New York: Raven Press, 1985. 308-310.
- Modifications developed by Newcomer Supply Laboratory.



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