

Part 4254A 10 Slide/Set

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Gram Positive & Gram Negative Bacteria, Animal Control Slides - Technical Memo

CONTROL SLIDES:

<u>.</u>

Part 4254B 98 Slide/Set

PRODUCT SPECIFICATIONS:

Tissue: Positive staining animal organ.

Fixation: Formalin 10%, Phosphate Buffered (Part 1090).

Section/Glass: Paraffin sections cut at 4 microns on Superfrost™ Plus slides.

Quality Control Stain: Brown-Brenn quality control stained slide(s) included.

Reactivity: Guaranteed product specific reactivity for one year from date of receipt. Revalidate after one year to verify continued reactivity.

Storage: 15-30°C in a light deprived and humidity controlled environment.

Intended Use: To verify histological techniques and reagent reactivity.

Before using unstained control slides, review the enclosed stained slide(s) to ensure that this tissue source is acceptable for testing needs.

CONTROL SLIDE VALIDATION:		
With Gram, Brown-Brenn Stain Kit:	Part 9123A	Individual Stain Solution
Solution A: Crystal Violet-Oxalate Stain, Alcoholic	250 ml	Part 10422
Solution B: Iodine, Gram, Aqueous	250 ml	Part 1140
Solution C: Acetone-Alcohol 1:1	250 ml	Part 10016
Solution D: Basic Fuchsin Stain 0.25%, Aqueous	250 ml	Part 1011
Solution E: Tartrazine Stain 0.25%. Acetic Aqueous	250 ml	Part 14016

APPLICATION:

Newcomer Supply Gram Positive & Gram Negative Bacteria, Animal Control Slides are for the positive histochemical staining of gram positive and gram negative bacteria in a naturally occurring infection.

PRESTAINING PREPARATION:

- 1. Heat dry sections in oven according to your laboratory protocol.
- 2. Filter Solution A: Crystal Violet-Oxalate Stain, Alcoholic.

STAINING PROCEDURE:

- Deparaffinize sections thoroughly in three changes of xylene, 3 minutes each. Hydrate through two changes each of 100% and 95% ethyl alcohols, 10 dips each. Wash well with distilled water.
 a. See Procedure Notes #1 and #2.
- 4. Stain in <u>freshly filtered</u> Solution A: Crystal Violet-Oxalate Stain, Alcoholic (Step #2) for 1 minute.
- 5. Rinse well in distilled water.
- 6. Mordant in Solution B: Iodine, Gram, Aqueous for 1 minute.
- 7. Rinse well in distilled water, removing excess iodine.
- 8. Decolorize in Solution C: Acetone-Alcohol 1:1 until blue stops running; 7-10 dips.
- 9. Rinse well in distilled water.
- 10. Place in Solution D: Basic Fuchsin Stain 0.25%, Aqueous for 90 seconds.
- 11. Rinse well in distilled water.
- 12. Dip once in Solution C: Acetone-Alcohol 1:1.
- 13. Counterstain in Solution E: Tartrazine Stain 0.25%, Acetic Aqueous for 5-15 seconds.
- 14. Rinse well in distilled water.
- Dehydrate in two changes of 100% ethyl alcohol, 5 dips each. Clear in three changes of xylene, 10 dips each; coverslip with compatible mounting medium.
 - a. Do not use 95% alcohol in the dehydration step.

RESULTS:

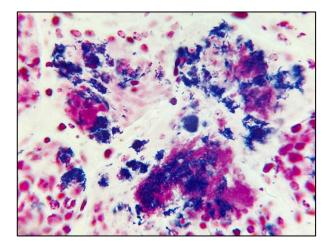
Gram positive bacteria Blue/violet Gram negative bacteria Red

PROCEDURE NOTES:

- 1. Drain slides after each step to prevent solution carry over.
- 2. Do not allow sections to dry out at any point during procedure.
- 3. If using a xylene substitute, follow manufacturer's recommendation for deparaffinization and clearing steps.

REFERENCES:

- Bancroft, John D., and Marilyn Gamble. *Theory and Practice of Histological Techniques*. 6th ed. Oxford: Churchill Livingstone Elsevier, 2008. 312-313.
- 2. Brown, J.H., and L. Brenn. "A Method for the Differential Staining of Gram Positive and Gram Negative Bacteria in Tissue Sections". *Bulletin of The Johns Hopkins* 48.2 (1931): 69-73.
- Luna, Lee G. Histopathologic Methods and Color Atlas of Special Stains and Tissue Artifacts. Gaitheresburg, MD: American Histolabs, 1992. 188-189.
- 4. Modifications developed by Newcomer Supply Laboratory.



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