

Part 4240 Revised January 2020

Giemsa Control Slides - Technical Memo

CONTROL SLIDES: Part 4240A Part 4240B 10 Slide/Set 98 Slide/Set

NEWCOMEI

PRODUCT SPECIFICATIONS:

Tissue: Positive staining spleen.

Fixation: Formalin 10%, Phosphate Buffered (Part 1090).

Section/Glass: Paraffin sections cut at 4 microns on Superfrost™ Plus slides.

Quality Control Stain: May-Grunwald Giemsa quality control stained slide(s) included.

Reactivity: Guaranteed product specific reactivity for one year from date of receipt. Revalidate after one year to verify continued reactivity.

Storage: 15-30°C in a light deprived and humidity controlled environment. **Intended Use:** To verify histological techniques and reagent reactivity.

Before using unstained control slides, review the enclosed stained slide(s) to ensure that this tissue source is acceptable for testing needs.

CONTROL SLIDE VALIDATION:

With May-Grunwald Giemsa Stain:

Jenner Stock Stain

Giemsa Stock Stain, Wolbach
Alcohol, Methanol Anhydrous, ACS
Acetic Acid 1%, Aqueous

Individual Stain Solution
Part 1210
Part 1121
Part 12236
Part 10012

APPLICATION:

Newcomer Supply Giemsa Control Slides are for the positive histochemical and differential staining of hematopoietic tissue.

NEWCOMER SUPPLY VALIDATION PROCEDURE:

- 1. Heat dry sections in oven according to your laboratory protocol.
- Deparaffinize sections thoroughly in three changes of xylene, 3 minutes each. Hydrate through two changes each of 100% and 95% ethyl alcohols, 10 dips each. Wash well with distilled water.
 - a. See Procedure Notes #1 and #2.
- Rinse in two changes of Alcohol, Methanol Anhydrous, ACS (12236); 3 minutes each.
- Prepare <u>fresh</u> Working Jenner Stain Solution just prior to use; combine and mix well.

a. Distilled Water 20 ml b. Jenner Stock Stain 20 ml

- 5. Place slides in <u>fresh</u> Working Jenner Stain Solution for 6 minutes.
- Prepare <u>fresh</u> Working Giemsa Stain Solution just prior to use; combine and mix well.

a. Distilled Waterb. Giemsa Stock Stain, Wolbach47 mlb. 3 ml

- Place slides directly into <u>fresh</u> Working Giemsa Stain Solution for 45 minutes.
- 8. Rinse quickly in distilled water.
- Differentiate <u>each slide individually</u> in Acetic Acid 1%, Aqueous (10012); 6-10 dips.
 - a. Check microscopically for well differentiated nuclei.
- 10. Rinse in distilled water.
- Dehydrate in two changes each of 95% and 100% ethyl alcohol. Clear in three changes of xylene, 10 dips each; coverslip with compatible mounting medium.

RESULTS:

Nuclei Blue/violet

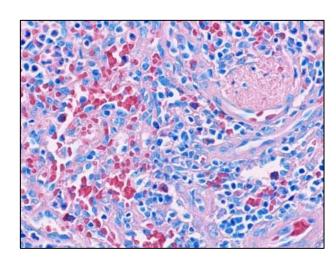
Cytoplasm Pink/rose to lighter blue shades

PROCEDURE NOTES:

- 1. Drain slides after each step to prevent solution carry over.
- 2. Do not allow sections to dry out at any point during procedure.
- The color range of the stained cells may vary depending upon fixation and degree of differentiation.
- If using a xylene substitute, closely follow the manufacturer's recommendations for deparaffinization and clearing steps.

REFERENCES:

- Luna, Lee G. Manual of Histologic Staining Methods of the Armed Forces Institute of Pathology. 3rd ed. New York: Blakiston Division, McGraw-Hill, 1968. 121-122.
- Shapiro, Stanley H., and Hilda Laufer. "Observations on Fixation and Staining of Bone Marrow Biopsies." The Journal of Histotechnology 11.3 (1988): 145-47.
- Sheehan, Dezna C., and Barbara B. Hrapchak. Theory and Practice of Histotechnology. 2nd ed. St. Louis: Mosby, 1980. 157.
- 4. Modifications developed by Newcomer Supply Laboratory.



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