

## Bile Control Slides – Technical Memo

<b>CONTROL SLIDES:</b>	<b>Part 4060A</b>	<b>Part 4060B</b>
	10 Slide/Set	98 Slide/Set

### PRODUCT SPECIFICATIONS:

**Tissue:** Positive staining liver.

**Fixation:** Formalin 10%, Phosphate Buffered (Part 1090).

**Section/Glass:** Paraffin sections cut at 4 microns on Superfrost™ Plus slides.

**Quality Control Stain:** Bile Stain, Hall's Method quality control stained slide(s) included.

**Reactivity:** Guaranteed product specific reactivity for one year from date of receipt. Revalidate after one year to verify continued reactivity.

**Storage:** 15-30°C in a light deprived and humidity controlled environment.

**Intended Use:** To verify histological techniques and reagent reactivity.

**Before using unstained control slides, review the enclosed stained slide(s) to ensure that this tissue source is acceptable for testing needs.**

### CONTROL SLIDE VALIDATION:

<b>With Bile Stain, Hall's Method:</b>	<b>Individual Stain Solution</b>
Fouchet Reagent	Part 1095
Van Gieson Stain	Part 1404

### APPLICATION:

Newcomer Supply Bile Control Slides are for the positive histochemical staining of bile (bilirubin) substances in tissue sections and to distinguish bile pigments from other tissue pigments.

### PRESTAINING PREPARATION:

1. Heat dry sections in oven according to your laboratory protocol.
2. Filter Fouchet Reagent with high quality filter paper prior to use.

### NEWCOMER SUPPLY VALIDATION PROCEDURE:

3. Deparaffinize sections thoroughly in three changes of xylene, 3 minutes each. Hydrate through two changes each of 100% and 95% ethyl alcohols, 10 dips each. Wash well with distilled water.
  - a. See Procedure Notes #1 and #2.
4. Place slides in freshly filtered Fouchet Reagent for 5 minutes.
5. Wash in three changes of tap water; rinse in distilled water.
6. Stain sections in Van Gieson Stain for 5 minutes.
7. Rinse slides quickly in 95% ethyl alcohol.
8. Dehydrate in two changes each of 95% and 100% ethyl alcohol. Clear in three changes of xylene, 10 dips each; coverslip with compatible mounting medium.

### RESULTS:

Bile	Emerald green to olive drab
Connective tissue	Pink to red
Background	Yellow

### PROCEDURE NOTES:

1. Drain staining slides after each step to prevent solution carry over.
2. Do not allow sections to dry out at any point during procedure.
3. If using a xylene substitute, closely follow the manufacturer's recommendations for deparaffinization and clearing steps.

### REFERENCES:

1. Carson, Freida L., and Christa Hladik. *Histotechnology: A Self-Instructional Text*. 3rd ed. Chicago, Ill.: American Society of Clinical Pathologists, 2009. 268-269.
2. Sheehan, Dezna C., and Barbara B. Hrapchak. *Theory and Practice of Histotechnology*. 2nd ed. St. Louis: Mosby, 1980. 219.
3. Modifications developed by Newcomer Supply Laboratory.

