Safranin O Stain for Cartilage - Technical Memo

SOLUTION:

<table>
<thead>
<tr>
<th>Solution</th>
<th>125 ml</th>
<th>500 ml</th>
</tr>
</thead>
<tbody>
<tr>
<td>Safranin O Stain 1%, Aqueous</td>
<td>Part 1350A</td>
<td>Part 1350B</td>
</tr>
</tbody>
</table>

Additionally Needed:

<table>
<thead>
<tr>
<th>Solution</th>
<th>125 ml</th>
<th>500 ml</th>
</tr>
</thead>
<tbody>
<tr>
<td>Xylene, ACS</td>
<td>Part 1445</td>
<td></td>
</tr>
<tr>
<td>Alcohol, Ethyl Denatured, 100%</td>
<td>Part 10841</td>
<td></td>
</tr>
<tr>
<td>Alcohol, Ethyl Denatured, 95%</td>
<td>Part 10842</td>
<td></td>
</tr>
<tr>
<td>Hematoxylin Stain Set, Weigert Iron</td>
<td>Part 1409</td>
<td></td>
</tr>
<tr>
<td>Fast Green Stain 2.5%, Aqueous</td>
<td>Part 10852</td>
<td></td>
</tr>
<tr>
<td>Acetic Acid 1%, Aqueous</td>
<td>Part 10012</td>
<td></td>
</tr>
</tbody>
</table>

APPLICATION:

Newcomer Supply Safranin O Stain for Cartilage procedure is used to stain cartilage, mucin, and mast cells granules in tissue sections and can assist in demonstrating articular cartilage loss in arthritic and other articular disease processes.

Safranin O is a basic dye that stains growth plate cartilage and articular cartilage (proteoglycans, chondrocytes and type II collagen) varying shades of red. The intensity of Safranin O staining is proportional to the proteoglycan content in the cartilage tissue. Fast Green counterstains the non-collagen sites and provides a clear contrast to the Safranin O staining.

METHOD:

Fixation: Formalin 10%, Phosphate Buffered (Part 1090)

Technique: Decalcified paraffin sections cut at 5 microns

Solutions: All solutions are manufactured by Newcomer Supply, Inc.

All Newcomer Supply stain procedures are designed to be used with Coplin jars filled to 40 ml following the staining procedure provided below.

STAINING PROCEDURE:

1. Deparaffinize sections thoroughly in two changes of xylene, 3 minutes each. Hydrate through two changes each of 100% and 95% ethyl alcohols, 10 dips each. Wash well with distilled water.
   a. See Procedure Notes #2 and #3.
2. Wash well in running tap water; rinse in distilled water.
3. Prepare fresh Weigert Iron Hematoxylin (1409); mix well.
   a. Solution A: Ferric Chloride, Aqueous 20 ml
   b. Solution B: Hematoxylin 1%, Alcoholic 20 ml
4. Stain slides in fresh Weigert Iron Hematoxylin for 10 minutes.
5. Wash in running tap water for 10 minutes; rinse in distilled water.
   a. See Procedure Note #4.
6. Prepare 0.25% Fast Green Stain, Aqueous; combine and mix well.
   a. Fast Green Stain 2.5%, Aqueous (10852) 5 ml
   b. Distilled Water 45 ml
7. Stain slides in 0.25% Fast Green Stain, Aqueous for 5 minutes.
8. Rinse directly in Acetic Acid 1%, Aqueous (10012); 10-15 seconds.
9. Place directly in Safranin O Stain 1%, Aqueous for 5 minutes.
10. Dehydrate in two changes each of 95% and 100% ethyl alcohol. Clear in three changes of xylene, 10 dips each; coverslip with compatible mounting medium.

RESULTS:

Cartilage: Red to orange
Mucin and mast cell granules: Red to orange
Bone, connective tissue and cytoplasm: Green
Nuclei: Black

PROCEDURE NOTES:

1. Certain fixatives and decalciying agents such as EDTA, nitric acid and hydrochloric acid can work to extract proteoglycans and result in weak Safranin O cartilage staining.
2. Drain staining rack/slides after each step to prevent solution carry over.
3. Do not allow sections to dry out at any point during staining procedure.
4. If Weigert Iron Hematoxylin is not completely washed from tissue sections, nuclear and cytoplasmic staining will be compromised.
5. If using a xylene substitute, closely follow the manufacturer's recommendations for deparaffinization and clearing steps.

REFERENCES:

5. Modifications developed by Newcomer Supply Laboratory.

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